



Penn Medicine | Hospital of the
University of Pennsylvania

3400 Spruce Street, Philadelphia, PA 19104

DIVISION OF LABORATORY MEDICINE

Michael Feldman, MD, PhD

Vice Chairman of Pathology for Clinical Services
6 Gates, Rm 6043 • Client Services: 215-662-4808

Vivianna Van Deerlin, MD, PhD

Molecular Pathology Laboratory Director

Molecular Pathology Laboratory • 7 Gates Bldg • (215) 662-6121

Label Area

Location

Contact Phone Number

Indication, clinical information, and relevant prior testing:

Date and Time of Collection

Contact FAX Number or email

Name of Ordering Physician

ICD10 code (required)

FOR FURTHER INFORMATION ON TESTS AND SPECIMEN REQUIREMENTS, PLEASE REFER TO <https://www.testmenu.com/HUP>

SPECIMEN TYPE (see back of form for details)

☐ Peripheral blood (EDTA)

☐ Bone marrow aspirate

☐ Cord blood

☐ Gynecologic specimen (ThinPrep)

☐ Nasopharyngeal swab

☐ Other (specify): _____

☐ Tissue: Site(s) _____ Case # _____

☐ Fresh ☐ Frozen ☐ Fixed: Specify fixative _____

☐ Buccal swab

☐ Fine needle aspirate: Site _____

☐ Bronchoalveolar lavage (BAL)

ONCOLOGY TESTING

RNA-based testing:

☐ *BCR-ABL1* RT-PCR (Qual to Qnt)

☐ *BCR-ABL1* RT-PCR (p210 Quantitative)

☐ *BCR-ABL1* RT-PCR (p190 Quantitative)

☐ *PML-RARA* RT-PCR (Qualitative)

☐ Leukemia translocation panel

☐ *FLT3* mutation analysis (ITD and D835)

☐ *JAK2* p.V617F mutation analysis

☐ IDH 1 Variant Analysis

☐ IDH 2 Variant Analysis

☐ *BRAF* mutation analysis (codon 600)

☐ *IGH* rearrangement

☐ TCR gamma (*TRG*) rearrangement

☐ *MGMT* methylation assay

GENETIC TESTING

☐ Cystic fibrosis (139 *CFTR* mutation panel):

☐ Screening ☐ Diagnosis (incl. CBAVD)

☐ Factor V Leiden (*F5*) mutation analysis

☐ Prothrombin (*F2*) mutation analysis

☐ *C9orf72* hexanucleotide repeat expansion

☐ *HTT Repeat Expansion Analysis*

IDENTITY TESTING

☐ Chimerism analysis (specify type and IDs)

☐ Pre-transplant evaluation (provide recipient buccal swab and blood)

Recipient name: _____

Donor 1 name or ID: _____

Donor 2 name or ID: _____

☐ Post-transplant evaluation

☐ Unfractionated ☐ Myeloid, CD33/66b

☐ T cell, CD3

☐ Graft versus host disease (contact lab for specimen types)

☐ Other identity testing (contact lab)

Specify: _____

INFECTIOUS DISEASE TESTING

☐ Respiratory virus panel (RVP)

☐ Viral load, specify: ☐ BKV ☐ EBV ☐ CMV ☐ HBV ☐ HIV ☐ HCV

☐ HCV genotyping (includes HCV viral load)

☐ HPV high-risk DNA, gynecologic (includes 16/18 genotyping)

OTHER TESTING

☐ Other, specify: _____

FOR LABORATORY USE ONLY

Requesting pathologist:

Resident/Fellow performing triage:

Non-MP accession number (if applicable):

Approved tests and comments:

FOR LAB USE
ONLY

AFFIX CERNER
LABEL

GENERAL GUIDE TO SPECIMEN TYPES FOR MOLECULAR PATHOLOGY TESTING

Refer to <https://www.testmenu.com/HUP> for more detailed specimen requirements by test.

Peripheral blood: EDTA containing blood tubes (pink or lavender) are appropriate for all genetic testing and for DNA and RNA-based oncology testing and identity testing where peripheral blood is the specimen of interest. RNA-based testing requires more volume, thus the pink top tube is the preferred tube as one lavender top tube does not provide sufficient volume. Peripheral blood is also appropriate for all viral load testing (pearl white PPT top preferred but pink top is acceptable). For post-transplant chimerism analysis unfractionated blood (pink preferred) will be tested along with specified cellular subsets (requires additional blood tubes for each subset). Lavender and pink top may be used interchangeably provided that sufficient volume is collected.

Tissue that is fresh (in Michel's medium) or frozen may be submitted for genetic testing as well as *IGH* and *TRG* rearrangement studies. Tissue fixed in formalin is acceptable for selected oncology testing including *IGH* and *TRG*, *BRAF* mutation analysis and *MGMT* methylation analysis.

Bone marrow aspirate in lavender tubes can be used for DNA and RNA-based oncology molecular testing and identity testing.

Cord blood is appropriate for pre-transplant chimerism analysis.

Buccal swabs are used for recipient pre-transplant chimerism analysis.

Gynecologic specimens in ThinPrep medium are appropriate for HPV DNA testing.

Fine needle aspirates are appropriate for oncologic molecular tests including *BRAF* mutation analysis.

Nasopharyngeal swabs (using NP flocked swab Lawson# 195443) are appropriate for respiratory virus panel (RVP) testing which includes influenza A/B, RSV A/B, parainfluenza virus 1/2/3 and 4 adenovirus, human metapneumovirus, human rhinovirus/enterovirus, coronavirus (NOT SARS-CoV-2), chlamydia pneumoniae and mycoplasma pneumoniae.

Bronchoalveolar lavage (BAL) specimens can be used for RVP testing and CMV viral load.

Please call the laboratory if in doubt about the acceptability of any specimen or specimen type.
Molecular Pathology Laboratory Main Number: (215) 662-6121.